

## I. Arrays in VNMR

Vnmr allows you to quickly define experiments in which a series of spectra can be obtained as a function of any NMR parameter. For example, you can obtain an array of spectra where transmitter frequency, pulse width, temperature, decoupling state, sweep width, etc., are varied. You can define arrays of one dimension by varying a single parameter. Alternatively, you can define multi-dimensional arrays in which two or more parameters are varied (on the Gemini-300 you are limited to arrays of 3 parameters).

You can array any parameter by simply typing the command `array` and answering the questions about the desired parameter and the size and range of the array. You can also define an array by manually setting the parameter to the desired values. For example, to obtain a series of 10 spectra where each successive spectra is acquired with twice the number of scans as the previous one, you would type `nt=1,2,4,8,16,32,64,128,256,512,1024`. To obtain spectra with the decoupler turned on and off, you would enter `dm='yyy','nnn'`. **NOTE:** on the Gemini-300, you need to specify whether the parameter you are varying is array 1, 2, or 3. Do so by placing a 1, 2, or 3 in parenthesis after the parameter name, e.g. `NT(1)=2,4,6,8`, etc.

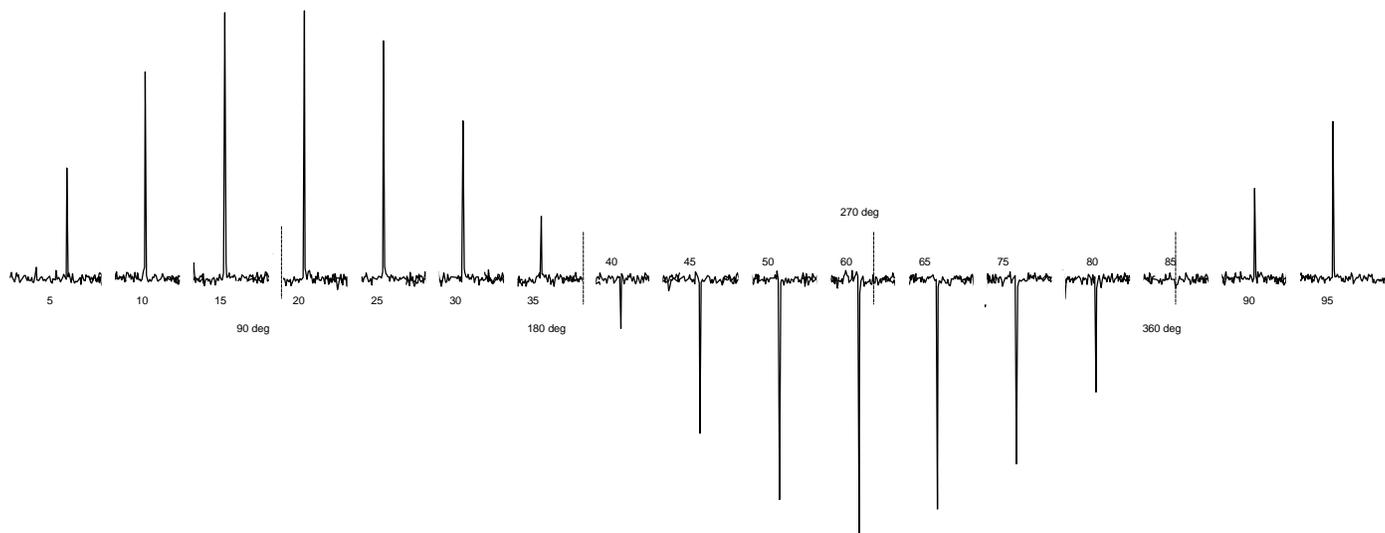
The following section describes how to perform several basic NMR measurements using the standard varian 2 pulse acquisition sequence `s2pul` and arrayed acquisition of spectral data.

## II. Simple Arrays using `s2pul`

### A. Measuring 90 degree pulse widths

You can measure the 90 degree pulse width of the selected nucleus by making an array `pw`. The following spectra was obtained on the Gemini-300 using the array `pw=5,10,15,20,25,30,35,40,45,50,55,60,65,70,75,80,85,90,95`. When measuring the pulse width, the parameters `p1` and `d2` are set to zero and `d1` is set to ca.  $5 \times T_1$ .

Figure 1. Peak Intensity vs. Pulse Width

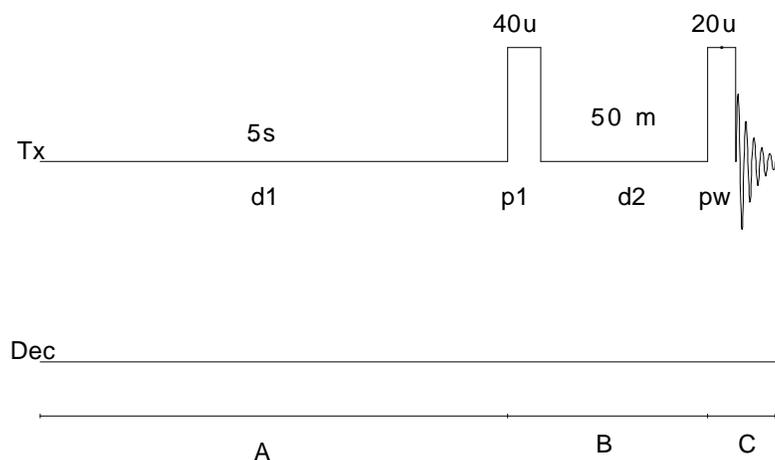




## B. Spin-Lattice Relaxation Times ( $T_1$ )

Arrays greatly simplify the measurement of spin-lattice relaxation times or  $T_1$ . The standard 2 pulse sequence (**s2pul**), in combination with arrays, can be used to perform  $T_1$  measurements using inversion-recovery sequence, as shown below. The inversion ( $180^\circ$ ) pulse is performed with **p1**, which is followed by the recovery delay **d2**. **d2** is arrayed to observe the relaxation of magnetization to equilibrium as a function of time.

**Figure 2. Inversion Recovery  $T_1$  Pulse Sequence Generated With s2pul.**



$T_1$  measurements require a long ( $5 \times T_1$ ) delay of time **d1**, followed by a  $180^\circ$  inversion pulse **p1**. The magnetization is allowed to recover for a time **d2**. The residual magnetization is then examined by a quantitative  $90^\circ$  pulse **pw**. The sequence is repeated for **nt** transients at various values of **d2**.

**d1** =  $5 \times T_1$

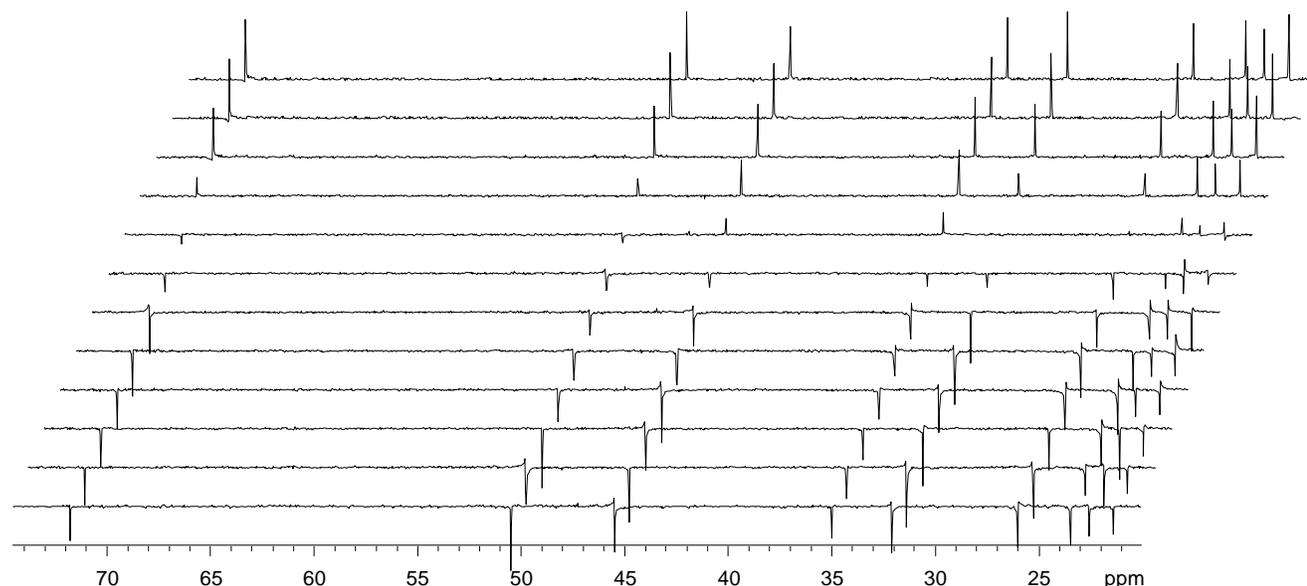
**p1** =  $180^\circ$

**d2** = array from ca.  $0.01 \times T_1$  to  $3-5 \times T_1$

**pw** =  $90^\circ$

An example of an arrayed series of spectra obtained using the inversion-recovery method is shown in Figure 3.  $T_1$  experiments can be easily acquired using **s2pul** and arrays. To acquire a  $T_1$  data set, obtain a simple 1D spectrum. Type **dot1** and answer the questions about minimum and maximum expected  $T_1$  values and the desired time for the experiment. The macro **dot1** will generate set **pw** to  $90^\circ$  and **p1** to  $180^\circ$ . **dot1** will set the equilibration delay **d1** to ca. 5 times the longest expected  $T_1$  value and **dot1** will then generate an array of relaxation delays **d2**. You can adjust **d1**, **d2** and **nt** to change the overall time required, if so desired (type **time** to find out how long the experiment will take. Finally, type **ega** to acquire the data.

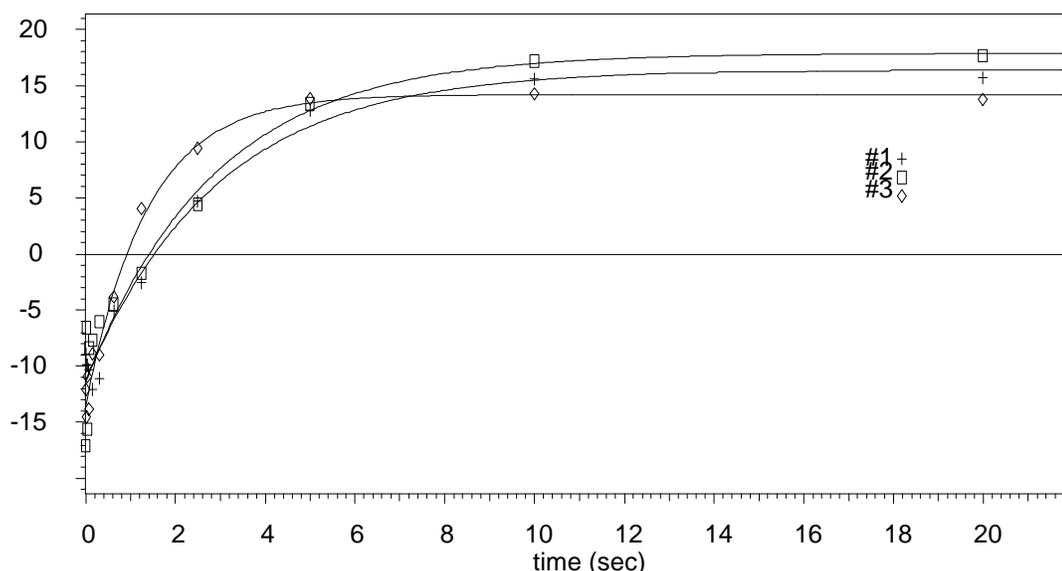
**Figure 3. Example of a Carbon  $T_1$  Measurement (Menthol).**





VNMR has an excellent set of tools for analyzing the results of the  $T_1$  experiment. To process the  $T_1$  data, display the last spectrum and set the threshold for peak picking. Type **ps(n)** where **n** is the number of the last spectrum. Phase the spectrum and set the threshold, in exactly the same way you would with a simple 1D spectrum. Adjust the display so that only six peaks are shown and type **dpf** (or **dll**). This will generate a list of peaks in the spectrum. Now type **fp**. The command **fp** will find the intensity of each of the peaks in the displayed region and will do so for all the spectra in the array. Type **t1** to calculate the  $T_1$  value for each of the displayed peaks. The command **t1** will generate a list of peak intensities for each peak, and the quality of the fit to the estimated  $T_1$  value. You can generate a graphical display of this data using the **expl** (Figure 4). The command **expl** will display the fit of the data. The following graph was generated by displaying three of the peaks in the  $T_1$  example shown earlier, followed by **dpf**, **fp**, **t1**, and **expl**. The exponential analysis can be output to the plotter by typing **pexpl page expl** and **pexpl** accept the peak numbers as options. For example, if you had displayed six peaks and analyzed the  $T_1$  values with **fp**, followed by **t1**, you could generate the  $T_1$  curves for peaks 2 and 5 (of the six) by typing **expl(2,5)**.

Figure 4. Example of  $T_1$  Analysis.



### C. Kinetics and Variable Temperature Experiments

Kinetics and Variable Temperature experiments are straightforward to set up using arrays. VNMR has a parameter called the pre-acquisition delay, **pad**. The spectrometer will wait **pad** seconds before acquiring the spectrum. For a kinetics measurement, simply array **pad** to leave the desired time between successive spectra. Alternatively, a local macro program called **kineticset** can be used to automatically define an array of **pad** values that increase in user-definable stages that take into consideration the exponential rate of change in kinetic experiments.

Once you have defined the array of **pad** values, type **ga** to acquire the spectra. VNMR contains tools to analyze the spectra in a manner that is very similar to that of  $T_1$  data. First, display the last spectrum in the kinetics series and set the threshold for peak picking. Type **ps(n)** where **n** is the number of the last spectrum. Phase the spectrum and set the threshold, in exactly the same way you would with a simple 1D spectrum. Adjust the display so that only six peaks are shown and type **dpf** (or **dll**) to generate the list of peaks in the displayed portion of the spectrum. Next type **fp** to determine the intensity of each of the peaks in the displayed region of the array. Type **kin** or **kind** to fit the data for each peak to an increasing or decreasing exponential function, respectively. The

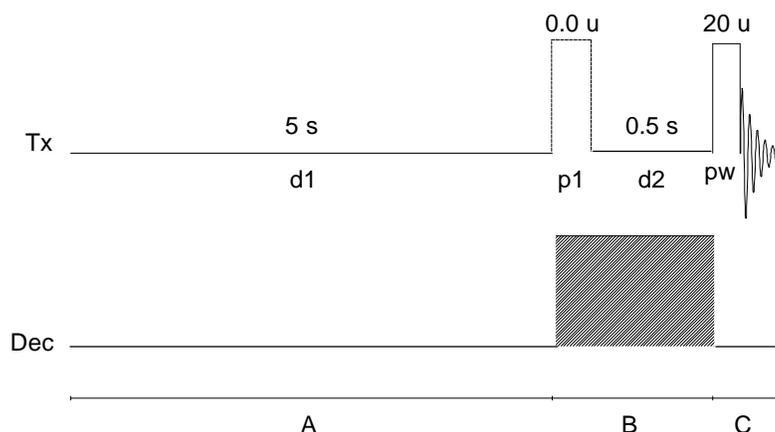


commands **kini** and **kind** generate a list of peak intensities for each peak, and give errors to the exponential fit. As with  $T_1$  data analysis, you can generate a graphical display of this data using the **expl** as shown in Figure 4.

## D. Single Frequency Decoupling Measurements

In progress

## E. Homonuclear One-Dimensional NOE Measurements



In the homonuclear NOE experiment a spectrum is acquired with single-frequency (selective) CW irradiation of one resonance. This spectrum is subtracted from a second spectra acquired without irradiation of the peak.

**d1=5 x  $T_1$  (typically 5-20 s)**

**d2=1 to 5 x  $T_1$**

**pw=90° or less**

**dm='nyn'**

**dmm='ccc', dof=desired frequencies**

The homonuclear NOE measurement requires an array of decoupler frequencies. The first spectrum is acquired with the decoupler off-resonance from any peaks. Successive spectra are acquired with irradiation of a single peak for a time **d2**. The decoupler is turned off and the transient is acquired after the pulse **pw**. The spin-system is allowed to return to equilibrium during the time **d1**. Setup, acquisition, and processing of the homonuclear NOE measurement is covered in detail in a separate handout.

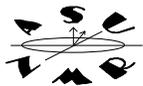
## IV. Interleaving Block Acquisition of Arrays Using *nt*, *il* and *bs*.

In progress

## IV. Useful Commands Associated with the *s2pul* Pulse Sequence

In progress.

<b>da</b>	
<b>expl(n,m,etc.),</b>	
<b>pexpl(n,m,etc.)</b>	
<b>fp</b>	
<b>t1</b>	
<b>kineticset</b>	



kini, kind  
pl(n,m,o), pl('all')  
dssa, dssh, dss  
vo, ho  
sd, sda  
pad  
time  
dot1  
clradd, spadd,  
    spsub, addi  
array